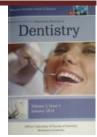


Comparing proliferative activity of Giant Cell Granuloma Lesions with Osteoclastic and Histiocytic derivation using PCNA,TRAP and CD163



### Ghaly A.M.\*, Gaballah E.T.\*\*, Elnagdy S.Y.\*\*\*, Farag D.A.\*\*\*\*

\*Demonstratoer of oral pathology, faculty of dentistry, Mansoura university.

\*\* Vice Dean for post graduate and research Affairs, Delta university for science and technology, Professor of Oral pathology, Faculty of dentistry, Mansoura University.

\*\*\* Head and professor of oral pathology department, Faculty of dentistry, Mansoura University.

\*\*\*\* Lecturer of Oral pathology, Faculty of dentistry, Mansoura University.

#### Abstract:

**Background:** Giant cell granulomas of the jaws are lesions that arise either peripherally in periodontal ligament and mucoperiosteum or centrally in the bone. The aim of this study was to evaluate expression of tartrate-resistant acid phosphatase (TRAP) and CD163 proteins in multinucleated giant cells and mononuclear cells and compare proliferative activity between central giant cell granuloma (CGCG) and peripheral giant cell granuloma (PGCG) by immunohistochemistry of PCNA.

**Methods:** The study was conducted on 40 formalin fixed paraffin embedded tissue blocks (20 CGCG ,20 PGCG). Five micron thick sections were cut from the paraffin blocks; two for hematoxylin and eosin to confirm diagnosis, while three sections were prepared for the immunohistochemical detection of TRAP, CD163 and PCNA by using *Avidin-Biotin complex according to the manufacturer's instructures*.

**Results:** There was statistically significant difference regarding CD163 between CGCG and PGCG, while there was no statistically significant differences of TRAP and PCNA between CGCG and PGCG. Also, there was high negative correlation between immunoexpression of TRAP and CD163 in CGCG and PGCG. There was no correlation between TRAP and PCNA and between CD163 and PCNA.

**Conclusions:** From the findings of the present study, it can be concluded that the histogenesis of MNGs of CGCG and PGCG was suggested to be of the osteoclastic origin, while there was no differences between proliferative activity between these two lesions.

### Introduction

The giant cell granuloma of oral cavity and jaws are lesions that occur peripherally from periodontal ligament and periosteum (PGCG) or centrally in the bone(CGCG). Histologically, both peripheral and

central giant cell granulomas are characterized by presence of few to many multinucleated giant cells in a fibrocellular connective tissue stroma<sup>(1)</sup>, although multi-nuclear giant cells are the hallmark of these lesions, the <u>histogenesis</u> of the giant cells has not been specified yet <sup>(2)</sup>.

Tartrate-resistant acid phosphatase (TRAP) is an ironcontaining enzyme that is found in humans. It occurs in diverse tissues including bone and cartilage .TRAP is highly expressed in osteoclasts as well as chondroclasts therefore, used as a specific histochemical marker for these cells. TRAP promots the dephosphorylation of bone matrix phosphoproteins like osteopontin and bone sialoprotein<sup>(3)</sup>, so it's related to bone resorption<sup>(1)</sup>.

Cluster of differentiation163 (CD163) antigen is a member of the scavenger receptor cysteine-rich (SRCR) super family class B, it's expressed by cells of macrophages lineage. The function of CD163 is a homeostatic one and related to the binding of Hemoglobin-Haptoglobin complexes<sup>(4)</sup>.

Proliferating cell nuclear antigen (PCNA) was originally characterised as a DNA sliding clamp for replicative DNA polymerases and used as a marker of cell proliferation<sup>(5)</sup>. Therefore, the present study was carried out to compare the histogenesis of MNGCs in CGCG and PGCG using immunohistochemical identification of TRAP and CD163.

And to compare proliferative activity between these lesions by using of immunohistochemistry of PCNA.

#### Material and Methods

The present study was conducted on 40 formalin fixed paraffin embedded tissue blocks (20 CGCG and 20 PGCG). Five micron thick sections were cut from each paraffin block; two were stained with hematoxylin and eosin to confirm diagnosis, while three sections were prepared for the immunohistochemical detection of TRAP, CD163 and PCNA by using Avidin-Biotin complex according to the manufacturer's instructures. The immunoreactivity of TRAP, CD163 and PCNA was evaluated by computer assisted digital image analysis (Digital morphometric study)

### Results

In CGCG, giant cells reacted for TRAP marker with a mean number of positive cells (88.86±SD6.023) and were negative to CD163 marker. In mononuclear cells, expression of TRAP with a mean number of positive cells was (14.63±SD 2.45) and for CD163 was (13.37±SD3.43) (table1,2) (figure1,3), while the mean number of positive cells to PCNA was (78.75±SD23.34) (table3) (figure5).

In PGCG, giant cells reacted for TRAP marker with a mean number of positive cells ( $86.81\pm$ SD 8.41) and were negative to CD163 marker. In mononuclear cells, expression of TRAP with a mean number of positive cells was ( $14.05\pm$ SD 3.001) and for CD163 was ( $9.76\pm$ SD2.34). (table1,2) (figure2,4), while the mean number of positive cells to PCNA was ( $73.58\pm$ SD18.98)(table3) (figure6).

_		CGCG	PGCG	P
TRAP (%)	Giant cells	Mean	88.86	86.810.44
		±SD	6.023	8.411
	Mononuclear cells	Mean	14.63	14.050.57
		±SD	2.450	3.001

 $Table(1): Comparison \ of \ TRAP \ expression \ in \ CGCG \ \ and \ PGCG \ \ groups \ .$ 

### Table(2):Comparison of CD163 expression in CGCG and PGCG groups.

			CGCG	PGCG	Р
	Ciant colla	Mean	0	0	-
CD1(2(0/)	Giant cells	±SD	0	0	
CD163(%)	Mononuclear cells	Mean	13.37	9.767	0.002*
		±SD	3.432	2.337	

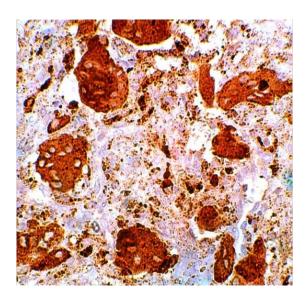
\*:significance <0.05

Test used: Student' t-test

## Table(3):Comparison of PCNA average area between CGCG & PGCG groups.

		CGCG	PGCG	Р
	Mean	2607	2498	0.7
PCNA(average area)	±SD	677.3	833.6	

\*:significance <0.05



multinucleated giant cells (red arrow) and mononuclear cells (green arrow) (TRAP X 400).

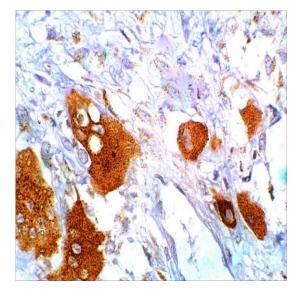
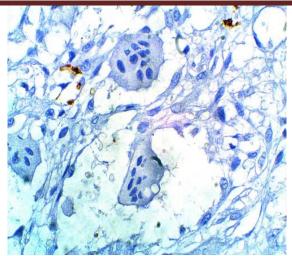
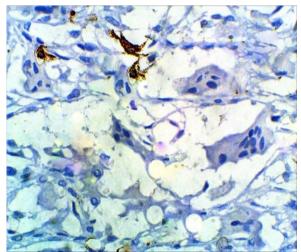


Figure 2: A case of PGCG with diffuse brown granular reaction in the

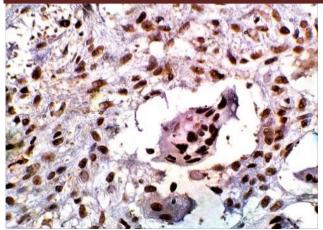
cytoplasm of the multinucleated giant cells (red arrow) and mononuclear cells (green arrow) (TRAP X 400).



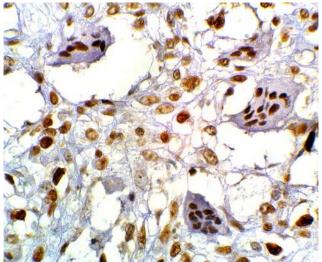
Fig(3):Acase of CGCG showing negative immunoreaction in the MNGCs (CD163X400).



Fig(4): Acase of PGCG showing negative immunoreaction in the MNGCs and positive mononuclear cells (CD163X400).



Fig(5): A case of CGCG showing prominent nuclear reaction to anti-PCNA in MNGC and fibroblasts (PCNAX400).



Fig(6): A case of PGCG showing prominent nuclear reaction to anti-PCNA in MNGC and fibroblasts (PCNAX400).

### Discussion

Microscopic examination of central and peripheral giant cell granuloma consists of multinuclear giant cells in a background of ovoid, spindle-shaped mesenchymal cells and foci of hemorrhage, although multi-nuclear giant cells are the hallmark of these lesions, the <u>histogenesis</u> of the giant cells has not been specified yet <sup>(2)</sup>.Some investigators believe that the giant cells show the immunohistochemical characteristics of <u>osteoclasts</u><sup>(6)</sup> while others have suggested the macrophages as an origin for these cells, it was also suggested that the stromal <u>mononuclear cells</u> play an important role in the evolution of giant cells.<sup>(7)</sup>

The present study was carried out to examine TRAP, CD163 and PCNA immunohistochemical expression in CGCG and PGCG to investigate the histogenesis of multinucleated giant cells and compare proliferative activity between central and peripheral giant cell granuloma lesions.

In the present study, the immunohistochemical expression of TRAP in CGCG and PGCG appeared in most of

# Mansoura Journal of Dentistry 2020;7(25):52-56

multinucleated giant with no statistical significant differences which was in accordance with <u>Itonaga</u> I. et al<sup>(8)</sup>, Flanagan AM <sup>(9)</sup>and Liu B<sup>(10)</sup> who suggested the osteoclastic phenotype of MNGCs, while <u>Torabinia</u> N et al <sup>(1)</sup> found high percentage of TRAP expression in 99% of CGCG and PGCG cases.

Similar to the findings of the present study, <u>Liu</u> B and coworkers<sup>(11)</sup> supposed that MNGCs in CGCG and PGCG of the jaws showed some similarities to the osteoclasts, such as the presence of acid phosphatase (TRAP) and osteoclast specific cellular antigens. But, their relation to the osteoclasts has never been fully established because the previous enzymes and antibodies were not unique or sufficient to identify the osteoclasts. Especially these antibodies did not confirm whether they had the potential to resorb the bone. In addition, previous studies only focused on the CGCG and PGCG, while MNGCs in other types of the giant cell containing lesions, such as cherubism and aneurysmal bone cyst, have rarely been investigated, although these lesions share considerable similarities in histomorphology.

Meanwhile, the findings of the present study showed that a group of mononuclear cells of stroma of CGCG and PGCG expressed osteoclastic TRAP marker with no statistical significant differences which was in accordance with Torabinia N et al <sup>(1)</sup> and Adkins et al<sup>(12)</sup> who suggested that the mononuclear cells considered as a progenitor of giant cells which were formed by fusion and adhesion of stromal mononuclear cells, but the underlying mechanism remains unknown.

On the contrary to the findings of the present study, N Mohtasham et al<sup>(13)</sup>, Kumar et al<sup>(14)</sup>, HJ Sherlin et al<sup>(15)</sup> and <u>Hallikeri K</u> et al<sup>(16)</sup> supposed a macrophage origin for MNGCs of CGCG and PGCG lesions.

In the present study, the immunohistochemical expression of CD163 in CGCG and PGCG appreared in most of mononuclear macrophages while negative in multinucleated giant cells which was in accordance to Kahn and co-workers<sup>(17)</sup> who stated that expression of CD163 is restricted to cells of macrophages lineage predominant in perivascular location due to metabolism of Hb is a main function of tissue macrophages because of their ability to engulf senescent erythrocytes (extravasated hemolysis) or take up Hb released from ruptured erythrocytes (intravascular hemolysis). Giant cells closely located to these extravasation areas<sup>(18)</sup>.

The findings of the current study showed a statistical significant differences in macrophages CD163 expression between CGCG ( $13.37\pm$ SD3.43) and PGCG ( $9.76\pm$ SD2.33) which was in accordance with Kumar et al <sup>(14)</sup> who stated that the number of macrophages were significantly higher in CGCG than PGCG.

In another study carried out by Leek et al., on breast carcinomas showed increased number of macrophages as a predictor of poor prognosis indicating that it plays a role in belligerent behaviour of tumour<sup>(19)</sup>. Similarly Lu CF et al., correlated poor prognosis with a high frequency of

macrophages in oral squamous cell carcinoma<sup>(20)</sup>. Similar to these findings, the present study revealed that macrophages were higher in CGCG than PGCG which suggest that it may

be considered as a predictor of poor prognosis signifying its aggressive behaviour.

On the contrary to the findings of the present study Pedreira and co-workers<sup>(21)</sup>revealed that MNGCs expressed CD163 ( $2.7 \pm$ SD 0.7) in oral paracoccidioidomycosis granulomas which explained by CD163 positive macrophages represent the main inflammatory cells in the immune response against fungus Paracoccidioides brasiliensis (Pb).

In the present study, the immunohistochemical expression of PCNA appeared in a fraction of the spindle shaped mononuclear cells and MNGCs with no statistical significant differences were reported between percentage of positive PCNA cells in central and peripheral giant cell granuloma which was in accordance with <u>Bo Liu</u> and co-workers<sup>(11)</sup>, <u>Itonaga</u> et al<sup>(8)</sup> and Houpis et al<sup>(22)</sup> who suggested that these cells were the proliferating tumor cells responsible for the biologic activity of the lesions.

On the contrary to the findings of the present study, Souza *et*  $al.^{(23)}$  concluded in their study that PCNA positive cells were more in PGCG. Thus, according to Souza *et al.* although CGCG is more aggressive, however, PGCG is more proliferative than CGCG.

The findings of the present work showed that MNGCs were strongly positive to anti-TRAP (osteoclastic marker) and negative to anti-CD163 (macrophage marker), this proved osteoclastic origin of MNGCs in central and peripheral giant cell granulomas. Meanwhile there was no statistical significant differences of the proliferative activity of central and peripheral giant cell granuloma regarding PCNA expression.

### References

1. Torabinia N, Razavi S, Shokrolahi ZJJoOP, Medicine. A comparative immunohistochemical evaluation of CD68 and TRAP protein expression in central and peripheral giant cell granulomas of the jaws. 2011;40(4):334-7.

2. Patil KP, Kalele KP, Kanakdande VD. Peripheral giant cell granuloma: a comprehensive review of an ambiguous lesion. Journal of the International Clinical Dental Research Organization. 2014;6(2):118.

3. Hayman AR, Cox TM. Tartrate-resistant acid phosphatase knockout mice. J Bone Miner Res. 2003;18(10):1905-7.

4. Fabriek BO, Dijkstra CD, van den Berg TKJI. The macrophage scavenger receptor CD163. 2005;210(2-4):153-60.

5. Maga G HU. Proliferation cell nuclear antigen (PCNA): a dancer with many partners. Journal of Cell Science. 2003:3051–60.

6. Khiavi MM, Aghbali AA, Halimi M, Kouhsoltani M, Hamishehkar HJJoo, JOMFP mp. Immunohistochemical expression of Src protein in peripheral and central giant cell granulomas of the jaws. 2013;17(3):358.

7. Varsha V, Hallikeri K, Girish H, Murgod SJJoo, JOMFP mp. Expression of CD34 and CD68 in peripheral giant cell granuloma and central giant cell granuloma: An immunohistochemical analysis. 2014;18(3):341.

8. Itonaga I, Hussein I, Kudo O, Sabokbar A, Watt-Smith S, Ferguson D, et al. Cellular mechanisms of osteoclast formation and lacunar resorption in giant cell granuloma of the jaw. Journal of oral pathology & medicine. 2003;32(4):224-31.

9. Flanagan AM, Tinkler SM, Horton M, Williams D, Chambers TJC. The multinucleate cells in giant cell granulomas of the jaw are osteoclasts. 1988;62(6):1139-45.

10. Liu B, Yu SF, Li TJJJoop, medicine. Multinucleated giant cells in various forms of giant cell containing lesions of the jaws express features of osteoclasts. 2003;32(6):367-75.

11. Liu B, Yu SF, Li TJ. Multinucleated giant cells in various forms of giant cell containing lesions of the jaws express features of osteoclasts. Journal of oral pathology & medicine. 2003;32(6):367-75.

 Medicine, Oral Pathology. Cellular morphology and relationships in giant-cell lesions of the jaws. 1969;28(2):216-22.
 Mohtasham N, Saghravanian N, Fatemi B, Vahedi M, Afzal-Aghaee M, Kadeh HJBjoo. A comparative study of osteopontin and MMP-2 protein expression in peripheral and central giant cell granuloma of the jaws. 2017.

14. Kumar VV, Krishanappa SJ, Prakash SG, Channabasaviah GH, Murgod S, Pujari R, et al. Quantification and correlation of angiogenesis with macrophages by histomorphometric method in central and peripheral giant cell granuloma: an immunohistochemical analysis. 2016;10(3):ZC01.

15. Kumar A, Sherlin HJ, Ramani P, Natesan A, Premkumar PJIJoDR. Expression of CD 68, CD 45 and human leukocyte antigen-DR in central and peripheral giant cell granuloma, giant cell tumor of long bones, and tuberculous granuloma: An immunohistochemical study. 2015;26(3):295.

16. Hallikeri K, Acharya S, Koneru A, Trivedi DJJJoAC, Insights R. Evaluation of microvessel density in central and peripheral giant cell granulomas. 2015;2(1):20-5.

 Kahn A, Chaushu G, Ginene L, Vered MJJoO, Surgery M. Age and Expression of CD163 and Colony-Stimulating Factor 1 Receptor (CD115) Are Associated With the Biological Behavior of Central Giant Cell Granuloma. 2017;75(7):1414-24.
 Pogrel AMJAoms. The diagnosis and management of giant cell lesions of the jaws. 2012;2(2):102.

19. Leek RD, Lewis CE, Whitehouse R, Greenall M, Clarke J, Harris ALJCr. Association of macrophage infiltration with angiogenesis and prognosis in invasive breast carcinoma. 1996;56(20):4625-9.

20. Lu CF, Huang CS, Tjiu JW, Chiang CPJH, Sciences NJft, Head Sot, et al. Infiltrating macrophage count: a significant predictor for the progression and prognosis of oral squamous cell carcinomas in Taiwan. 2010;32(1):18-25.

21. Pedreira RdPG, de Carli ML, Beijo LA, Nonogaki S, Pereira AAC, Junior NVR, et al. Oral paracoccidioidomycosis granulomas are predominantly populated by CD163+ multinucleated giant cells. 2016;181(9-10):709-16.

22. Houpis CH, Tosios KI, Papavasileiou D, Christopoulos PG, Koutlas IG, Sklavounou A, et al. Parathyroid hormone–related peptide (PTHrP), parathyroid hormone/parathyroid hormone–related peptide receptor 1 (PTHR1), and MSX1 protein are expressed in central and peripheral giant cell granulomas of the jaws. 2010;109(3):415-24.

23. Souza P, Mesquita R, Gomez RJOd. Evaluation of p53, PCNA, Ki-67, MDM2 and AgNOR in oral peripheral and central giant cell lesions. 2000;6(1):35-9.